

Review

Biochemical and medical importance of vanadium compounds

Jan Korbecki¹, Irena Baranowska-Bosiacka¹, Izabela Gutowska² and Dariusz Chlubek¹

¹Department of Biochemistry and Medical Chemistry, Pomeranian Medical University, Szczecin, Poland; ²Department of Biochemistry and Human Nutrition, Pomeranian Medical University, Szczecin, Poland

Vanadium belongs to the group of transition metals and is present in the air and soil contaminants in large urban agglomerations due to combustion of fossil fuels. It forms numerous inorganic compounds (vanadyl sulfate, sodium metavanadate, sodium orthovanadate, vanadium pentoxide) as well as complexes with organic compounds (BMOV, BEOV, METVAN). Depending on the research model, vanadium compounds exhibit antitumor or carcinogenic properties. Vanadium compounds generate ROS as a result of Fenton's reaction or of the reaction with atmospheric oxygen. They inactivate the Cdc25B, phosphatase and lead to degradation of Cdc25C, which induces G₂/M phase arrest. In cells, vanadium compounds activate numerous signaling pathways and transcription factors, including PI3K-PKB/Akt-mTOR, NF-KB, MEK1/2-ERK, that cause cell survival or increased expression and release of VEGF. Vanadium compounds inhibit p53-dependent apoptosis and promote entry into the S phase of cells containing functional p53 protein. In addition, vanadium compounds, in particular organic derivatives, have insulin-mimetic and antidiabetic properties. Vanadium compounds lower blood glucose levels in animals and in clinical trials. They also inhibit the activity of protein tyrosine phosphatase 1B. By activating the PI3K-PKB/Akt pathway, vanadium compaunds increase the cellular uptake of glucose by the GLUT4 transporter. The PKB/Akt pathway is also used to inactivate glycogen synthase kinase-3. The impact of vanadium compounds on inflammatory reactions has not been fully studied. Vanadium pentoxide causes expression of COX-2 and the release of proinflammatory cytokines in a human lung fibroblast model. Other vanadium compounds activate NF-κB in macrophages by activating IKKβ.

Key words: vanadium, pollution, cancer, diabetes, insulin-mimetic action, inflammation

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NATURAL OCCURRENCE OF VANADIUM

Vanadium is a transition metal, owing its name to Vanadís – Norse goddess of beauty and fertility. It is estimated that more than 60 thousand tons of this element are emitted into the atmosphere each year as the result of human activities (mostly combustion of fossil fuels) (Aragón & Altamirano-Lozano, 2001). This is due to high vanadium concentrations in both crude oil (3–260 μ g/g) and hard coal (14–56 μ g/g). Atmospheric pollution with vanadium of natural origin is relatively low and estimated at several tons annually. The consequence of emission of large amounts of vanadium into the atmosphere is the relatively high concentration (20– 300 ng/m^3) of this element in the air of big cities, with values reaching up to 10 mg/m³ observed in the New York City and other large urban agglomerations (Aragón & Altamirano-Lozano, 2001; Lin et al., 2004). Soils in areas not subject to anthropogenic changes contain small amounts of vanadium, originating mostly from volcanic rocks (Połedniok & Buhl, 2003; Nadal et al., 2004). Industrial activities result in a significant increase in these levels, reaching 19.3 $\mu g/g$ of soil in the vicinity of a crude oil refinery in Catalonia (Nadal et al., 2004). Vanadium present in soil is accumulated in plants (Nadal et al., 2004; Marcano et al., 2006). Contamination with vanadium is also observed in water reservoirs: rivers, lakes and seas. Bottom sediments of the Persian Gulf contain vanadium at concentrations as high as 100 µg/g of dry sediment (Pourang et al., 2005). About 10% of groundwater samples from California and some other states of the USA contain vanadium in amounts exceeding 25 µg/ dm³ (Wright & Belitz, 2010). This is due to vanadium being washed out of water-bearing rocks (Wright & Belitz, 2010).

As evidenced by studies of vanadium levels in the hair of residents of different countries, Poland's population as a whole is not significantly exposed to high levels of vanadium. The measured value is of the order of $0.055 \ \mu g/g$, being three times lower than the value of 0.171 µg/g for residents of the U.S., Canada or India (Stefańska et al., 2005). Hair vanadium content in students in Białystok is even lower (0.038 μ g/g) due to a non-polluted environment (Stefańska et al., 2005). On the other hand, vanadium pollution is observed in the Upper Silesia region (Połedniok & Buhl, 2003). Industrial pollution of the Silesian regions combined with automobile exhaust fumes is transported by rivers into the sea and are deposited in bottom sediments of the rivers. Thus, the sediments in the Bay of Szczecin are highly polluted with vanadium and other elements originating

e-mail: dchlubek@sci.pum.edu.pl

Abbreviations: BEOV, bis(ethylmaltolato)oxovanadium(IV); BKOV, bis(kojato)oxovanadium(IV); BMOV, bis(maltolato)oxovanadium(IV); Cdc25B₂, cell division control/cycle 25 homolog B₂; Cdc25C, cell division control/cycle 25 homolog C; CksHs1, human cyclin dependent kinase subunit type 1; COX-2, cyclooxygenase 2; CXCL10, C-X-C motif chemokine 10; EGFR, epidermal growth factor receptor; ERK, extracellular signal-regulated kinase; E2F, Transcription factor E2F; GLUT4, Glucose transporter type 4; GSK3, glycogen synthase kinase-3; HIF-1a, hypoxia inducible factor 1a; IC₅₀, half maximal inhibitory concentration; IkBa, inhibitor of kB activity a; IKK β , IkB kinase subunit β ; IL-6 Interleukin-6; IL-8, Interleukin-8; MAPK, mitogen-activated protein kinase; MEK1/2, MAPK/ERK kinase 1 and 2; MIP-2, macrophage inflammatory protein-2; mTOR, mammalian target of rapamycin; NF-kB, nuclear factor kB; NF-AT, nuclear factor of activated T-cells; p38, protein 38; p53, protein 53; PI3K, phosphatidylinositol 3-kinase; PKB/Akt, protein kinase B; PTP-1B, protein tyrosine phosphatase 1B; pBA, retinoblastoma protein; ROS, reactive oxygen species; SSB, single-strand break; TNFa, Tumor necrosis factor a; VEGF, vascular endothelial growth factor.

from distant regions (Glasby *et al.*, 2004). Due to the river runoff, vanadium pollution of the Bay of Szczecin is comparable to the pollution of the Persian Gulf oilfields (Glasby *et al.*, 2004; Pourang *et al.*, 2005).

VANADIUM IN LIVING ORGANISMS

After entering the circulatory system via the gastrointestinal or respiratory tract, vanadium compounds are transported by transferrin or, less commonly, by albumin or low-molecular components of plasma, such as citrates and, to a lesser extent, lactates or phosphates (Kiss et al., 2000). Next, vanadium compounds are accumulated in kidneys and, to a smaller degree, in spleen, bones and liver (Hansen et al., 1982). Human body contains ca. 100 µg of vanadium, with equilibrium between the amount of vanadium excreted from the body and the amount of vanadium absorbed from the outside environment (up to several dozen micrograms daily) (Byrne & Kosta, 1978; Kordowiak & Holko, 2009). For certain mammals, such as rats, vanadium is a necessary microelement; however, due to the omnipresence of this element at low concentrations, no necessity of nutritional intake of vanadium was determined in humans (Lin et al., 2004; Kordowiak & Holko, 2009).

Aquatic organisms, such as ascidians, accumulate vanadium in circulatory system cells known as vanadocytes (Kawakami *et al.*, 2006; Kawakami *et al.*, 2009). Blood vanadium levels in these organisms exceed 10 mM, while the sea concentration of vanadium is about 35 nM (Kawakami *et al.*, 2009). Vanadium compounds are transported into the cytoplasm of vanadocytes, bound and reduced to the +4 oxidation state by the binding proteins – vanabins, and finally deposited in the acidic environment of vacuoles as vanadium compounds in the +3 oxidation state (Kawakami *et al.*, 2006).

In human body, vanadium has an oxidation state of +4 or +5 (Kordowiak & Holko, 2009). Vanadium compounds in the +5 oxidation state (metavanadates or orthovanadates, forming oligomers) enter cells via anionic channels, while vanadium compounds in the +4 oxidation state (vanadyl cations) permeate the cellular membrane by diffusion (Fig. 1) (Aureliano & Gândara, 2005; Kordowiak & Holko, 2009). Vanadium forms numerous derivatives with low-molecular organic compounds. Vanadium organic derivatives have been synthesized since 1990s (Thomson *et al.*, 2009). Examples of such compounds include maltol complexes such as BMOV or

BEOV – compounds of insulin-mimetic activity characterized by low toxicity compared to inorganic vanadium compounds; naglivan, a cystein derivative complex, or BKOV (Fig. 2) (Scior *et al.*, 2009; Thompson *et al.*, 2009; Kordowiak & Holko, 2009). As the result of reactions with intracellular antioxidants, vanadium within the cells has predominantly an oxidation state of +4 (Aureliano & Gândara, 2005; Kordowiak & Holko, 2009).

Vanadium compounds in the +4 oxidation state are oxidized by atmospheric oxygen to the +5 oxidation state with accompanying emission of a superoxide anion radical (Cuesta et al., 2011). As a result of reduction with NADPH, the reaction may proceed with generation of hydrogen peroxide (Cuesta et al., 2011). Moreover, vanadium in the +4 oxidation state can be oxidized to the +5 oxidation state with generation of a hydroxyl radical via a Fenton-like reaction (Cuesta et al., 2011). In cells vanadium compounds oxidized to the +5 oxidation state by atmospferic oxygen or ROS are in equilibrium with vanadium compounds reduced to the +4 oxidation state by intracellular antioxidants. Thanks to their structural similarity to phosphate anions, orthovanadium anions may act as inhibitors of protein phosphatases or bind to such molecules as ADP or NAD to form ADPV and NADV, respectively (Crans et al., 2004).

VANADIUM COMPOUNDS AND DIABETES

Vanadium compounds have insulin-mimetic properties. First reports on therapeutic properties of vanadium compounds in diabetes appeared as early as in 1899 (Thompson & Orvig 2006). Many studies were conducted on inorganic and organic vanadium derivatives in induced-diabetes animal models, in which the studied compounds were found to impact the levels of glucose, cholesterol and triacylglycerols, with no harmful side effects upon prolonged administration (Yanardag et al., 2003; Koyuturk et al., 2005; Niu et al., 2007; Wei et al., 2007; Li et al., 2009). The studied vanadium compounds normalize renal function and the indicator liver enzyme levels in diabetic model animals (Yanardag et al., 2003; Koyuturk et al., 2005). Many experiments were also performed in diabetic patients, confirming the therapeutic effect of the studied vanadium compounds on blood glucose levels with little side effects (Thompson et al., 2009).

Vanadium compounds are characterized by multiple ways of action resulting in reduction of blood glucose







Figure 2. Examples of organic vanadium derivatives of medical importance According to Dong *et al.*, 2000; D'Cruz & Uckun, 2001; Scior *et al.*, 2008; Thompson *et al.*, 2009, (modified).

levels (Vardatsikos *et al.*, 2009). Thanks to their structural similarity to orthophosphate anions, the orthovanadate anion and vanadium organic derivatives are inhibitors of protein phosphotyrosine phosphatases (Fig. 4). (Crans *et al.*, 2004). They may inhibit the activity of PTP-1B,

which is an enzyme responsible for dephosphorylation of insulin receptors, causing insulin resistance (Scior et al., 2009; Scior et al., 2010). However, the mechanism of phosphotyrosine phosphatase inactivation is not yet fully understood, as it appears that this process may also be caused by free radicals (Bartosz, 2003). One may assume that vanadium compounds cause PTP-1B inhibition via ROS (Kaltschmidt et al., 2000). Another mechanism of reduction of blood glucose levels by vanadium compounds is the activation of PKB/Akt leading to increased uptake of glucose by the GLUT4 transporter (Vardatsikos et al., 2009). Activation of PKB/Akt results also in phosphorylation and inactivation of GSK3, leading to stimulation of the synthesis of glycogen from glucose (Vardatsikos et al., 2009).

VANADIUM AND TUMOR CELLS

In chemically-induced tumor models in experimental animals, vanadium compounds show chemopreventive properties by means of optimization of phase I and phase II xenobiotic transformation enzymes

(Bishayee et al., 2000; Ray et al., 2007; Chakraborty et al., 2007). Inorganic and organic vanadium compounds were tested in human tumor cell line models. The results were promising with respect to introduction of va-



Figure 3. Some antitumor and cancerogenic pathways of inorganic vanadium compounds According to Chen et al., 1999; Woo et al., 1999; Gao et al., 2002; Lapenna et al., 2002; Zhang et al., 2003; Zhang et al., 2004; Wozniak & Blasiak, 2004; Soares et al., 2008; Zhao et al., 2010; Morita et al., 2010; Parrondo, 2010 (modified).



Figure 4. Phosphatase 1B with its inhibitor, the orthovanadate anion. From left to right: model of the spatial structure of the enzyme; tertiary structure of the enzyme with the orthovanadate anion and aminoacyl residues in its catalytic center; a close-up of the catalytic center of the enzyme with a model of the orthovanadate anion and aminoacyl residues interacting with the inhibitor according to Brandão *et al.*, 2010 (modified).

nadium compounds into the therapy due to their low IC₅₀ (several micromoles depending on cell line and vanadium compound), antiproliferative and proapoptotic effects (Kordowiak et al., 2007; Holko et al., 2008; Fu et al., 2008; Klein et al., 2008; Molinuevo et al., 2008). Vanadium compounds are genotoxic and cause selective oxidation of pyrimidine bases and SSB-type DNA damages in tumor cells, which are characterized by less efficient DNA repair processes (Fig. 3) (Wozniak & Blasiak, 2004; Rodríguez-Mercado et al., 2011). Another mechanism of action of vanadium compounds on tumor cells is the opening of mitochondrial permeability transition pores, leading to the release of cytochrome c and induction of apoptosis (Soares et al., 2008; Zhao et al., 2010). Vanadium compounds inhibit the activity of phosphatase Cdc25B₂, responsible for dephosphorylation and activation of Cdk2 in cyclin-A and -B complexes (Woo et al., 1999). Inhibition of Cdc25B₂ induces \hat{G}_2/M phase arrest. Another mechanism of action of vanadium compounds on the G_2/M phase arrest is the degradation of $\hat{C}dc25C$ via MAPK cascades: ERK and p38 (Zhang et al., 2003; Liu et al., 2012). Another target for the vanadium compounds is CksHs1 (Arvai et al., 1995). Experiments on embryonic p53-knockout fibroblasts led to conclusion that vanadium compounds promote S phase entry of cells with wild-type p53 and induce G₂/M phase arrest of p53-knockout cells (Zhang et al., 2002). In addition, vanadium compounds activate NF-xB by the action of ROS in various cell types (Chen et al., 1999; Jaspers et al., 2000). The role of NF-xB in tumor cells is subject to discussion, as activation or overexpression of this transcription factor in non-tumor cells as well as in certain tumor cell lines leads to cell survival and inhibition of apoptosis (Parrondo, 2010). However, in prostate cancer cell lines treated with anticancer drugs, chemical activation of NF-xB increases the percentage of apoptotic cells (Parrondo, 2010). Inactivation of NF-xB in cells treated with vanadium compounds, such as pervanadate, decreases apoptotic cell death (Kaltschmidt et al., 2000).

One group of organic vanadium compounds tested as antitumor drugs includes complexes of the vanadyl cation with phenanthroline derivatives, such as METVAN [bis(4,7-dimethyl-l,10-phenanthroline) sulfatooxovanadium(IV); VO(SO₄)(Me₂-Phen)₂] (Narla *et al.*, 2000; Narla *et al.*, 2001). These vanadium compounds are characterized by antiproliferation IC₅₀ values of the order of several micromoles in many tumor cell lines and, when present at low concentrations, induce apoptosis and inhibit the cell cycle (Narla *et al.*, 2000; Dong *et al.*, 2000). METVAN is cytotoxic against many tumor cell lines (IC₅₀ of less than 1 μ M), reduces the invasiveness of leukemia by inhibiting the activity of metalloproteinases and damages mitochondria by generating ROS, thus causing apoptosis (Dong *et al.*, 2000; Narla *et al.*, 2001). One of side effects of the treatment is inhibition of spermatogenesis and apoptosis of male germ cells (D'Cruz & Uckun, 2001).

Vanadium compounds have carcinogenic properties and stimulate tumor development. Vanadyl cations and V₂O₅, a vanadium compound found in air pollution, generate ROS that cause DNA damage which may lead to mutations and, as a consequence, development of tumor cells (Ehrlich et al., 2008). Vanadium compounds have also antiapoptotic properties (Morita et al., 2010). Activation of NF-xB by ROS generated by vanadium compounds leads to cell survival and inhibition of apoptosis (Chen et al., 1999; Jaspers et al., 2000). In addition, by means of altered p53 phosphorylation, vanadates cause disturbances in the course of apoptosis (Suzuki et al., 2007; Morita et al., 2010). Another mechanism of carcinogenic action of vanadium compounds is activation of PIJK by generation of hydrogen peroxide (Gao et al., 2002). PI3K activates PKB/Akt, which promotes S phase entry via the E2F-pRb pathway (Zhang et al., 2004). In addition, activation of the PI3K-PKB/ Akt-mTOR pathway results in increased expression of HIF-1 α and, as a consequence, expression and release of VEGF (Gao et al., 2002). Expression of VEGF stimulated by vanadium compounds is controlled not only by the PI3K-PKB/Akt pathway, but also by two other pathways: MEK1/2-ERK or increase in intracellular calcium levels (Li et al., 2005). Release of VEGF causes angiogenesis and thus contributes to the development of tumor.

The effects of vanadium compounds depend on many factors, mainly on the type of cells, the type of vanadium compound and its dose. It appears that the proapoptotic or antiapoptotic effect of vanadium compounds depends largely on the cell type. The key protein, defects of which diametrically change the effects of vanadium compounds, is p53 (a large number of tumor cell types have defects in the gene encoding this protein). In p53defective cells (tumor cells or non-tumor p53-knock out cells), vanadium compounds inhibit the cell cycle and thus induce apoptosis (Zhang et al., 2002). Activation of NF-xB by ROS generated by vanadium compounds enhances the apoptotic effect (Parrondo, 2010). În contrast, in p53-functional cells, disturbed phosphorylation of p53 leads to inhibition of apoptosis (Morita et al., 2010). In addition, vanadium compounds stimulate the

cell cycle, thus inhibiting apoptosis, as both processes are mutually connected (Zhang et al., 2002). Moreover, NF-xB activation inhibits apoptosis of tumor cells. Another important fact is that vanadium compounds cause much more DNA damage in tumor cells compared to non-tumor cells when present at the same levels (Wozniak & Blasiak, 2004). Extensive DNA damage leads to apoptosis of tumor cells while a less intensive damage evoked by vanadium compounds in non-tumor cells may stimulate synthesis and activation of repair enzymes, thus protecting those cells from apoptosis. The above processes promote tumor cell growth at early stages of the disease and have an antitumor effect in the advanced stages of cancer. Studies in animals treated with carcinogens suggest that vanadium compounds used at low levels have selective effects on the tumor cells (Ray et al., 2007; Chakraborty et al., 2007).

VANADIUM COMPOUNDS AND INFLAMMATORY REACTIONS

The impact of vanadium compounds on inflammatory reactions has not been fully studied. Experiments conducted to date suggest that vanadates activate NF-xB, a transcription factor of key importance in inflammatory reactions (Chen et al., 1999; Ye et al., 1999). Studies conducted on RAW 264.7 macrophages showed that this was due to activation of IKK β and degradation of IzB α (Chen *et al.*, 1999; Ye *et al.*, 1999). Activation of NFxB leads to changes in expression of numerous genes, including TNFa and MIP-2, which belong to the CXC chemokine family (Ye et al., 1999; Chong et al., 2000). Another vanadium compound prevalent in air pollution and causing inflammatory reactions is vanadium pentoxide. Exposure to vanadium pentoxide-containing dust causes inflammatory reactions in lungs, leading to expression of, among others, COX-2, IL-6, IL-8 and CXCL10 (Ingram et al., 2007; Rondini et al., 2010). Vanadium pentoxide causes COX-2 expression in epithelial bronchial Beas-2B cells via the NF-AT pathway (Tang et al., 2007). Another pathway for the increase in COX-2 expression, encompassing EGFR and the p38 cascade, was observed in A249 lung cancer cells (Chen et al., 2006).

CONCLUSIONS AND FUTURE DIRECTIONS

Due to the ability to generate ROS, which exert nonspecific effects on different cell structures, vanadium compounds have many routes of action, sometimes diametrically opposite. They may have both antitumor and carcinogenic properties. The mechanisms of action of the vanadium compounds can be understood thanks to rapid advances in the knowledge of free radicals and the signaling pathways involving them. However, still little is known regarding the effect of vanadium compounds on the immune system and inflammatory reactions. New findings in this area may shed new light on the biochemical processes taking place in organisms treated with vanadium compounds. Currently, promising clinical trials of organic vanadium derivatives in the treatment of diabetes are under way. Soon they should be of common use. However, the effects of a long-term administration of low doses of vanadium as a potential carcinogen and correlation between the use of vanadium compounds and disorders of a free-radical background have not been fully studied yet. One of such diseases having a free-radical background is Parkinson's disease. Vanadium compounds induce ROS generation in the brain,

which may contribute to degeneration of dopaminergic neuronal cells of the substantia nigra, which in turn leads to Parkinson's disease (Afeseh Ngwa et al., 2009; Cuesta et al., 2011).

It is possible that an antitumor therapy using vanadium compounds will be developed in the near future. However, due to the carcinogenic effect of vanadium, such treatment should be combined with numerous other drugs (such as anti-VEGF antibodies) to enhance the therapeutic effect of vanadium.

REFERENCES

- Afeseh Ngwa H, Kanthasamy A, Anantharam V, Song C, Witte T, Houk R, Kanthasamy AG (2009) Vanadium induces dopaminergic neurotoxicity via protein kinase Cdelta dependent oxidative signaling mechanisms: relevance to etiopathogenesis of Parkinson's disease. Toxicol Appl Pharmacol 240: 273-285.
- Aragón AM, Altamirano-Lozano M (2001) Sperm and testicular modifications induced by subchronic treatments with vanadium (IV) in CD-1 mice. *Reprod Toxicol* **15**: 145–151.
- Arvai AS, Bourne Y, Hickey MJ, Tainer JA (1995) Crystal structure of the human cell cycle protein CksHs1: single domain fold with similarity to kinase N-lobe domain. J Mol Biol 249: 835-842. Aureliano M, Gândara RM (2005) Decavanadate effects in biological
- systems. J Inorg Biochem 99: 979–985. Bartosz G (2003) Obrazki z pola bitwy. In Druga twarz tlenu. pp 324– 325. PWN, Warszawa (in Polish).
- Bishayee A, Oinam S, Basu M, Chatterjee M (2000) Vanadium chemo-prevention of 7,12-dimethylbenz(a)anthracene-induced rat mammary carcinogenesis: probable involvement of representative hepatic phase I and II xenobiotic metabolizing enzymes. Breast Cancer Res Treat 63: 133–145.
- Brandão TA, Hengge AC, Johnson SJ (2010) Insights into the reaction of protein-tyrosine phosphatase 1B: crystal structures for transition state analogs of both catalytic steps. J Biol Chem 285: 15874–15883.
- Byrne AR, Kosta L (1978) Vanadium in foods and in human body fluids and tissues. Sci Total Environ 10: 17-30.
- Chakraborty T, Swamy AH, Chatterjee A, Rana B, Shyamsundar A, Chatterjee M (2007) Molecular basis of vanadium-mediated inhibi-
- Chatterjee M (2007) Molecular basis of Vanaduum-mediated inhibition of hepatocellular preneoplasia during experimental hepatocarcinogenesis in rats. J Cell Biochem 101: 244–258.
 Chen F, Demers LM, Vallyathan V, Ding M, Lu Y, Castranova V, Shi X (1999) Vanadate induction of NF-kappaB involves IkappaB kinase beta and SAPK/ERK kinase 1 in macrophages. J Biol Chem 274: 20307–20312.
- Chien PS, Mak OT, Huang HJ (2006) Induction of COX-2 protein expression by vanadate in A549 human lung carcinoma cell line through EGF receptor and p38 MAPK-mediated pathway. *Biochem* Biophys Res Commun 339: 562-568.
- Chong IW, Lin SR, Hwang JJ, Huang MS, Wang TH, Tsai MS, Hou JJ, Paulauskis JD (2000) Expression and regulation of macrophage inflammatory protein-2 gene by vanadium in mouse macrophages. Inflammation 24: 127–139.
- Crans DC, Smee JJ, Gaidamauskas E, Yang L (2004) The chemistry and biochemistry of vanadium and the biological activities exerted by vanadium compounds. Chem Rev 104: 849-902.
- Cuesta S, Francés D, García GB (2011) ROS formation and antioxidant status in brain areas of rats exposed to sodium metavanadate. Neurotoxicol Teratol 3: 1–6
- D'Cruz OJ, Uckun FM (2001) Bis(4,7-dimethyl and 5-dinitro-1,10phenanthroline) sulfato-oxovanadium(IV)-mediated in vivo male germ cell apoptosis. J Appl Toxicol 21: 331–339. Dong Y, Narla RK, Sudbeck E, Uckun FM (2000) Synthesis, X-ray
- structure, and anti-leukemic activity of oxovanadium(IV) complexes. J Inorg Biochem **78**: 321–330. Ehrlich VA, Nersesyan AK, Atefie K, Hoelzl C, Ferk F, Bichler J,
- Valic E, Schaffer A, Schulte-Hermann R, Fenech M, Wagner KH, Knasmüller S (2008) Inhalative exposure to vanadium pentoxide causes DNA damage in workers: results of a multiple end point
- study. Environ Health Perspect 116: 1689–1693.
 Fu Y, Wang Q, Yang XG, Yang XD, Wang K (2008) Vanadyl bisacetylacetonate induced G1/S cell cycle arrest via high-intensity ERK
- phosphorylation in HepG2 cells. J Biol Inorg Chem 13: 1001–1009. Gao N, Ding M, Zheng JZ, Zhang Z, Leonard SS, Liu KJ, Shi X, Jiang BH (2002) Vanadate-induced expression of hypoxia-inducible factor 1 alpha and vascular endothelial growth factor through phosphatidylinositol 3-kinase/Akt pathway and reactive oxygen species. *J Biol Chem* 277: 31963–31971.
- Glasby GP, Szefer P, Geldon J, Warzocha J (2004) Heavy-metal pollution of sediments from Szczecin Lagoon and the Gdansk Basin, Poland. Sci Total Environ 330: 249-69.

- Hansen TV, Aaseth J, Alexander J (1982) The effect of chelating agents on vanadium distribution in the rat body and on uptake by human erythrocytes. Arch Toxicol 50: 195-202
- Holko P, Ligeza J, Kisielewska J, Kordowiak AM, Klein A (2008) The effect of vanadyl sulphate ($VOSO_4$) on autocrine growth of human epithelial cancer cell lines. *Pol J Pathol* **59**: 3–8.
- Ingram JL, Antao-Menezes A, Turpin EA, Wallace DG, Mangum JB, Pluta LJ, Thomas RS, Bonner JC (2007) Genomic analysis of human lung fibroblasts exposed to vanadium pentoxide to identify candidate genes for occupational bronchitis. Respir Res 8: 34.
- Jaspers I, Samet JM, Erzurum S, Reed W (2000) Vanadium-induced kappaB-dependent transcription depends upon peroxide-induced actration of the p38 mitogen-activated protein kinase. Am J Respir Cell Mol Biol 23: 95–102.
- Kaltschmidt B, Kaltschmidt C, Hofmann TG, Hehner SP, Dröge W, Schmitz ML (2000) The pro- or anti-apoptotic function of NF-kappaB is determined by the nature of the apoptotic stimulus. Eur J Biochem 267: 3828-3835.
- Kawakami N, Ueki T, Amata Y, Kanamori K, Matsuo K, Gekko K, Michibata H (2009) A novel vanadium reductase, Vanabin2, forms a possible cascade involved in electron transfer. Biochim Biophys Acta 1794: 674-679.
- Kawakami N, Ueki T, Matsuo K, Gekko K, Michibata H (2006) Selective metal binding by Vanabin2 from the vanadium-rich ascidian, Ascidia sydneiensis samea. Biochim Biophys Acta **1760**: 1096–1101. Kiss T, Kiss E, Garribba E, Sakurai H (2000) Speciation of insulin-
- mimetic VO(IV)-containing drugs in blood serum. J Inorg Biochem 80: 65-73.
- Klein A, Holko P, Ligeza J, Kordowiak AM (2008) Sodium orthova-nadate affects growth of some human epithelial cancer cells (A549, HTB44, DU145). Folia Biol (Krakow) 56: 115-121
- Kordowiak AM, Holko P (2009) Pochodne wanadu jako związki o istotnym znaczeniu biologicznym. Część I. Działanie przeciwcukrzycowe. Post Biol Kom 36: 361-376 (in Polish)
- Kordowiak AM, Klein A, Goc A, Dabros W (2007) Comparison of the effect of VOSO4, Na3VO4 and NaVO3 on proliferation, viability and morphology of H35-19 rat hepatoma cell line. Pol J Pathol 58: 51 - 57
- Koyuturk M, Tunali S, Bolkent S, Yanardag R (2005) Effects of vanadyl sulfate on liver of streptozotocin-induced diabetic rats. Biol Trace Elem Res 104: 233-247.
- Lapenna D, Ciofani G, Bruno C, Pierdomenico SD, Giuliani L, Giamberardino MA, Cuccurullo F (2002) Vanadyl as a catalyst of human lipoprotein oxidation. Biochem Pharmacol 63: 375-380.
- Li J, Tong Q, Shi X, Costa M, Huang C (2005) ERKs activation and calcium signaling are both required for VEGF induction by vanadium in mouse epidermal Cl41 cells. Mol Cell Biochem 279: 25-33.
- Li M, Smee JJ, Ding W, Crans DC (2009) Anti-diabetic effects of sodium 4-amino-2,6-dipicolinatodioxovanadium(V) dihydrate in streptozotocin-induced diabetic rats. J Inorg Biochem 103: 585-589
- Lin TS, Chang CL, Shen FM (2004) Whole blood vanadium in Taiwanese college students. Bull Environ Contam Toxicol 73: 781-786.
- Liu TT, Liu YJ, Wang Q, Yang XG, Wang K (2012) Reactive-oxygen-species-mediated Cdc25C degradation results in differential antiproliferative activities of vanadate, tungstate, and molybdate in the PC-3 human prostate cancer cell line. J Biol Inorg Chem 17: 311–320. Marcano L, Carruyo I, Fernández Y, Montiel X, Torrealba Z (2006)
- Determination of vanadium accumulation in onion root cells (Allium cepa L.) and its correlation with toxicity. Biocell 30: 259-267
- Molinuevo MS, Cortizo AM, Etcheverry SB (2008) Vanadium(IV) complexes inhibit adhesion, migration and colony formation of UMR106 osteosarcoma cells. Cancer Chemother Pharmacol 61: 767-773.
- Morita A, Yamamoto S, Wang B, Tanaka K, Suzuki N, Aoki S, Ito A, Nanao T, Ohya S, Yoshino M, Zhu J, Enomoto A, Matsumoto Y, Funatsu O, Hosoi Y, Ikekita M (2010) Sodium orthovanadate inhibits p53-mediated apoptosis. Cancer Res 70: 257-265.
- Nadal M, Schuhmacher M, Domingo JL (2004) Metal pollution of soils and vegetation in an area with petrochemical industry. Sci Total Environ 321: 59-69.
- Narla RK, Chen CL, Dong Y, Uckun FM (2001) In vivo antitumor activity of bis(4,7-dimethyl-1,10-phenanthroline) sulfatooxovanadium (IV) (METVAN [VO(SO4)(Me2-Phen)2]). Clin Cancer Res 7: 2124-2133
- Narla RK, Dong Y, D'Cruz OJ, Navara C, Uckun FM (2000) Bis(4,7dimethyl-1,10-phenanthroline) sulfatooxovanadium (IV) as a novel apoptosis-inducing anticancer agent. Clin Cancer Res 6: 1546-1556.
- Narla RK, Dong Y, Klis D, Uckun FM (2001) Bis(4,7-dimethyl-1,10phenanthroline) sulfatooxovanadium (IV) as a novel antileukemic agent with matrix metalloproteinase inhibitory activity. Clin Cancer Res 7: 1094-1101.
- Narla RK, Dong Y, Uckun FM (2001) Apoptosis inducing novel anti-leukemic agent, bis(4,7-dimethyl-1,10 phenanthroline) sulfatooxovanadium(IV) [VO(SO4)(Me2,Phen)2] depolarizes mitochondrial membranes. Leuk Lymphoma 41: 625-634.
- Niu Y, Liu W, Tian C, Xie M, Gao L, Chen Z, Chen X, Li L (2007) Effects of bis(alpha-furancarboxylato)oxovanadium(IV) on glucose

metabolism in fat-fed/streptozotocin-diabetic rats. Eur J Pharmacol 572: 213-219.

- Parrondo R, de las Pozas A, Reiner T, Rai P, Perez-Stable C (2010) NF-kappaB activation enhances cell death by antimitotic drugs in human prostate cancer cells. Mol Cancer 9: 182-195.
- Połedniok J, Buhl F (2003) Speciation of vanadium in soil. Talanta 59:
- Pourang N, Nikouyan A, Dennis JH (2005) Trace element concentrations in fish, surficial sediments and water from northern part of the Persian Gulf. Environ Monit Assess 109: 293-316.
- Ray RS, Ghosh B, Rana A, Chatterjee M (2007) Suppression of cell proliferation, induction of apoptosis and cell cycle arrest: chemopreventive activity of vanadium in vivo and in vitro. Int | Cancer 120: 13-23.
- Rodríguez-Mercado JJ, Mateos-Nava RA, Altamirano-Lozano MA (2011) DNA damage induction in human cells exposed to vanadium oxides in vitro. Toxicol In Vitro 25: 1996-2002.
- Rondini EA, Walters DM, Bauer AK (2010) Vanadium pentoxide induces pulmonary inflammation and tumor promotion in a straindependent manner. Part Fibre Toxicol 7: 9.
- Scior T, Guevara-García JA, Melendez FJ, Abdallah HH, Do QT, Bernard P (2010) Chimeric design, synthesis, and biological assays of a new nonpeptide insulin-mimetic vanadium compound to inhibit protein tyrosine phosphatase 1B. Drug Des Devel Ther 4: 231-242.
- Broten (Joshe Phospitalae Phospitalae (2009) Antidiabetic Bis-Malto-lato-OxoVanadium(IV): conversion of inactive trans- to bioactive cis-BMOV for possible binding to target PTP-1B. Drug Des Devel Ther 2: 221-231
- Soares SS, Henao F, Aureliano M, Gutiérrez-Merino C (2008) Vanadate induces necrotic death in neonatal rat cardiomyocytes through mitochondrial membrane depolarization. Chem Res Toxicol 21: 607-618
- Stefańska E, Ostrowska L, Czapska D, Karczewski J, Borawska M (2005) Hair vanadium content and nutritional status of students of the Medical University of Białystok. Rocz Panstw Zakl Hig 56: 157-163.
- Suzuki K, Inageda K, Nishitai G, Matsuoka M (2007) Phosphorylation of p53 at serine 15 in A549 pulmonary epithelial cells exposed to vanadate: involvement of ATM pathway. Taxicol Appl Pharmacol 220: 83-91.
- Tang H, Sun Y, Xiu Q, Lu H, Han H (2007) Cyclooxygenase-2 induction requires activation of nuclear factor of activated T-cells in Beas-2B cells after vanadium exposure and plays an anti-apoptotic role. Arch Biochem Biophys 468: 92-99.
- Thompson KH, Lichter J, LeBel C, Scaife MC, McNeill JH, Orvig C (2009) Vanadium treatment of type 2 diabetes: a view to the future. J Inorg Biochem 103: 554-558.
- Thompson KH, Orvig C (2006) Vanadium in diabetes: 100 years from Phase 0 to Phase I. J Inorg Biochem 100: 1925–1935.
- Vardatsikos G, Mehdi MZ, Šrivastava AK (2009) Bis(maltolato)-oxovanadium (IV)-induced phosphorylation of PKB, GSK-3 and FOXO1 contributes to its glucoregulatory responses (review). Int J Mol Med 24: 303-309.
- Wei D, Li M, Ding W (2007) Effect of vanadate on gene expression of Wei D, Li W, Ding W (2007) Effect of variatine of gene capitosian of the insulin signaling pathway in skeletal muscle of streptozotocin-induced diabetic rats. J Biol Inorg Chem 12: 1265–1273.
 Woo ES, Rice RL, Lazo JS (1999) Cell cycle dependent subcellular distribution of Cdc25B subtypes. Oncogene 18: 2770–2776.
 Wozniak K, Blasiak J (2004) Vanadyl sulfate can differentially damage Control of the Arth Train(2007) 20: 74-66.
- DNA in human lymphocytes and HeLa cells. Arch Toxicol 78: 7-15. Wright MT, Belitz K (2010) Factors controlling the regional distribu-
- tion of vanadium in groundwater. Ground Water 48: 515-525. Yanardag R, Bolkent S, Karabulut-Bulan O, Tunali S (2003) Effects of
- vanadyl sulfate on kidney in experimental diabetes. Biol Trace Elem Res 95: 73-85.
- Ye J, Ding M, Zhang X, Rojanasakul Y, Nedospasov S, Vallyathan V, Castranova V, Shi X (1999) Induction of TNFalpha in macrophages by vanadate is dependent on activation of transcription factor NFkappaB and free radical reactions. Mol Cell Biochem 198: 193-200.
- Zhang Z, Chen F, Huang C, Shi X (2002) Vanadate induces G2/M phase arrest in p53-deficient mouse embryo fibroblasts. J Environ Pathol Toxicol Oncol 21: 223–231.
- Zhang Z, Gao N, He H, Huang C, Luo J, Shi X (2004) Vanadate activated Akt and promoted S phase entry. *Mol Cell Biochem* 255: 227-237
- Zhang Z, Leonard SS, Huang C, Vallyathan V, Castranova V, Shi X (2003) Role of reactive oxygen species and MAPKs in vanadateinduced G2/M phase arrest. Free Radic Biol Med 34: 1333-1342.
- Zhao Y, Ye L, Liu H, Xia Q, Zhang Y, Yang X, Wang K (2010) Vanadium compounds induced mitochondria permeability transition pore (PTP) opening related to oxidative stress. J Inorg Biochem 104: 371-378.