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Review

# Physiology and pathophysiology of vascular signaling controlled by guanosine 3',5'-cyclic monophosphate-dependent protein kinase<sup>\*</sup>

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Recent medical advances suggest that the cellular natriuretic peptide/cGMP and NO/cGMP effector systems represent important signal transduction pathways especially in the cardiovascular system. These pathways also appear to be very interesting targets for the possible prevention of cardiovascular diseases. Exciting candidates for prevention include cGMP-dependent signaling networks initiated by natriuretic peptides (NP) and nitric oxide (NO) which are currently explored for their diagnostic and therapeutic potential. cGMP signaling contributes to the function and interaction of several vascular cell types, and its dysfunction is involved in the progression of major cardiovascular diseases such as atherosclerosis, hypertension and diabetic complications. This review will take a focussed look at key elements of the cGMP signaling cascade in vascular tissue. Recent advances in our knowledge of cGMP-dependent protein kinases (cGK, also known as PKG), the potential for assessing the functional status of cGMP signaling and the possible cross talk with insulin signaling will be reviewed.

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Abbreviations: sGC, soluble guanylate cyclases; cGK, cGMP-dependent protein kinase; MLC, myosin light chain; NOS, NO synthase; PDE, phosphodiesterase; SNP, sodium nitroprusside; VASP, vasodilator-stimulated phosphoprotein; VSMC, vascular smooth muscle cell.

Abnormalities in endothelial and vascular smooth muscle cell function, as well as pathological activation of human platelets, play a decisive role in atherosclerosis and its complications (Hu *et al.*, 2002; Bhatt & Topol, 2003). These factors contribute to the pathogenesis of cardiovascular and cerebrovascular diseases which remain the major cause of death in most industrialized countries. Substantial efforts were undertaken in the last decades to elucidate underlying mechanisms, and some progress has been made.

Thirty years ago Schultz et al. (1977) and Katsuki et al. (1977) described the stimulation of soluble guanylate cyclases (sGC) by both NO and organic nitrates. The discovery of NO as "endothelium-derived relaxing factor" (EDRF) that regulates vascular tone was a major advance in this field (Furchgott & Zawadzki, 1980; Ignarro et al., 1987; Palmer et al., 1987). NO is synthesized by NO-synthases (endothelial eNOS, inducible iNOS, neuronal nNOS) from L-arginine. NO induces activity of the target enzyme sGC, which results in increasing tissue levels of the second messenger cGMP. cGMP activates cGMP- dependent protein kinase (cGK) that promotes vasorelaxation via phosphorylation of proteins that regulate intracellular  $Ca^{2+}$  levels. In addition to these effects on vascular smooth muscle, the NO/cGMP/PKG pathway affects various heart functions, platelets, immune cells and neurotransmission (von der Leyen & Dzau, 2001). Therefore, in essentially all mammalian cells this pathway is fundamental and highly regulated. Current research focuses on various steps of this signaling pathway with the aim to use this knowledge for better diagnostics and therapeutic applications.

## cGMP/cGK

Two pathways regulate the synthesis of cGMP. In the first one, cGMP is generated from GTP in response to different natriuretic

peptides (NP), namely the atrial (ANP), B-type (BNP) or C-type (CNP) natriuretic peptides. In the second pathway, cGMP is synthesized by soluble guanylyl cyclase in response to NO as mentioned above.

Two mammalian genes and three isotypes of cGK have been described so far:

- type I cGK consists of an  $\alpha$  and a  $\beta$  isoform, splice variants of a single gene. It is also identified as the most prominent cGK isotype in the cardiovascular system. Very high levels of cGKI are found in vascular smooth muscle cells, endothelial cells and platelets.
- type II cGK is mainly expressed in intestine, kidney and brain.

cGK signaling plays an important role in vascular biology, in regulating smooth muscle tone and in proliferation and differentiation of vascular smooth muscle cell (VSMC) (Ruth, 1999; Lincoln et al., 2001). In addition, it also regulates endothelial cell (i.e. permeability and motility) and platelet function (Lohmann et al., 1997; Ruth, 1999; Munzel et al., 2003). Further insight into the function of the two protein kinases has been obtained by the analysis of their substrates and by the generation of mice deficient in either cGK I or II. Homozygous deletion of the cGKI gene in mice abolishes NO/cGMP-dependent relaxation of vascular (Pfeifer et al., 1998), visceral (Pfeifer et al., 1998; Ny et al., 2000) and penile smooth muscle (Hedlund et al., 2000), resulting in severe vascular and intestinal dysfunction with death at an early age (Pfeifer et al., 1998). Additionally, the knockout mice showed impaired NO/cGMP-dependent inhibition of platelet activation (Massberg et al., 1999).

The phenotype of cGKII knockout mice included normal lifespan (Pfeifer *et al.*, 1996), decreased longitudinal bone growth (Pfeifer *et al.*, 1996), decreased intestinal chloride secretion (Pfeifer *et al.*, 1996; Vaandrager *et al.*, 2000) and altered renin secretion (Wagner *et al.*, 1998).

Results obtained with knockout mice discussed here support that cGKI plays an important role in the cardiovascular system. It also seems that cGKII has an influence on haemodynamic parameters *via* regulation of renin release and ion transport in the kidney.

cGK in different cell types has numerous effects. In the next section we discuss two of the best characterized pathways and examples of NO/cGMP/cGK signaling.

# ROLE OF cGMP SIGNALING PATHWAY IN VSMC

The serine/threonine protein kinase cGK mediates VSMC relaxation by catalyzing the phosphorylation of specific substrate proteins (Munzel et al., 2003). Many of the cGK substrate proteins, that have been implicated in relaxation, are involved in reducing intracellular Ca<sup>2+</sup> concentration or lowering the Ca<sup>2+</sup> sensitization. Both effects result in inhibition of myosin light chain (MLC) phosphorylation and therefore cause smooth muscle cells (SMC) contraction (Ammendola et al., 2001; Lincoln et al., 2001; Pfitzer. 2001). Reduced intracellular Ca<sup>2+</sup> concentration occurs due to various actions like Ca<sup>2+</sup> release from intracellular stores, Ca<sup>2+</sup> uptake mediated by calcium adenosine 5'-trisphosphatase of the endoplasmatic reticulum (ER) and Ca<sup>2+</sup> influx. Phosphorylation of IRAG (InsP<sub>3</sub>R associated cGMP kinase substrate; InsP<sub>3</sub>R: InsP<sub>3</sub> receptor; InsP<sub>3</sub>: inositol 1,4,5-triphosphate) (Schlossmann et al., 2000) by cGKI results in inhibition of IP<sub>3</sub>/IRAG-mediated  $Ca^{2+}$  release (Ammendola *et al.*, 2001) from the ER and therefore in activation of MLC kinase and vasoconstriction. Ca<sup>2+</sup> uptake is normally mediated by calcium ATPase which could be inhibited by cGKI-dependent phosphorylation of phospholamban. cGKI also phosphorylates a BK<sub>Ca</sub> channel, which leads to  $\mathrm{K}^+$ efflux from the cell, hyperpolarization, inhibition of Ca<sup>2+</sup> entry through voltage-dependent ion channels and relaxation (Fukao et al., 1999; Hofmann et al., 2000; Lincoln et al., 2001).

In summary, a number of cGKI-dependent mechanisms have been identified that regulate the cytosolic  $Ca^{2+}$  concentration. However, the precise physiological *in vivo* significance of these different pathways is still debated.

cGKI apparently also affects smooth muscle tone by reducing the Ca<sup>2+</sup> sensitivity of the contractile apparatus (Ruth, 1999). cGK phosphorylation of RhoA at Ser 188 induces RhoA translocation from the membrane to the cytosol whereby RhoA is inactivated. Thus it inhibits Rho kinase and consequently activates MLC phosphatase to dephosphorylate MLC. Consequently, cGKI-mediated activation of MLC phosphatase and calcium desensitization might involve inhibition of Rho/Rho kinase-dependent and -independent pathways (Sauzeau *et al.*, 2000; Etter *et al.*, 2001).

Other studies have suggested that NO and cGMP inhibit VSMC proliferation, a hallmark of many vascular disorders (Young *et al.*, 2000).

# ROLE OF cGMP SIGNALING PATHWAY IN PLATELETS

Over the past decade the role of platelets evolved from passive participants in the coagulation cascade to that of active synthesizer of humoral factors that potentiate both clot formation and inflammation. Antiplatelet therapy is presently the golden standard in the secondary prevention of cardiovascular incidents (Bhatt & Topol, 2003). Dipyridamole – an enhancer of NO/cGMP/cGK signaling pathway – is one of the very effective drugs (in combination with aspirin) in the secondary prevention of ischemic strokes again demonstrating the importance of this pathway for platelet activation/aggregation.

In vivo, platelets are continually exposed to the endothelial-derived factors NO and prostacyclin (PG-I<sub>2</sub>), which inhibit and limit unwarranted platelet activation by increasing the level of intracellular cAMP and cGMP (Nolte *et al.*, 1994; Schwarz *et al.*, 2001) (Fig. 1). Elevation of cGMP, induced by NO or pharmacological drugs such as the antihyper-

are also known to moderately raise platelet cGMP levels and possibly to limit and counteract the extent of platelet activation. However, recent published data suggest a

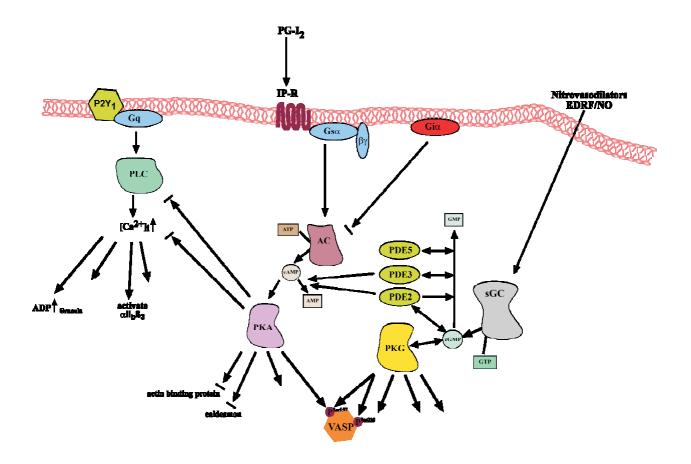


Figure 1. NO/cGMP/cGK-mediated mechanisms for platelet activation.

cGKI affects diverse signaling pathways leading to platelet activation. Abbreviations: AC, adenylate cyclase; EDRF, endothelium-derived relaxing factor; IP-R, prostaglandin receptor; PG-I<sub>2</sub>, prostaglandin I<sub>2</sub>; PKA, cAMP-dependent protein kinase; PKG, cGMP-dependent protein kinase (cGK); PLC, phospholipase C; sGC, soluble guanylate cyclase; VASP, vasodilator-stimulated phosphoprotein.

tensive, NO-releasing agent sodium nitroprusside (SNP) and the cGMP-phosphodiesterase type 5 (PDE 5), has been recognized to cause inhibition of platelets (Bhatt & Topol, 2003; Aktas *et al.*, 2003). One crucial effect is again the activation of cGK and the subsequent phosphorylation of specific target proteins such as the vasodilator-stimulated phosphoprotein (VASP).

Agonists which stimulate platelets such as thrombin, ADP, thromboxane receptor agonists and collagen (Haslam *et al.*, 1978) stimulatory role of cGMP and cGK in von Willebrand factor (vWF)- and thrombin-induced platelet activation. Li *et al.* (2003a, 2003b) report a stimulatory effect on MAP kinases and the fibrinogen receptor by activating cGK. Two groups, S. Watson's group and our own group, could not confirm these results. Our group (Gambaryan *et al.*, 2004) demonstrated a potent cGK-unrelated inhibition of human platelet activation by both cGK activators and inhibitors, using the conditions published by Li *et al.* (2003a, 2003b). Further evidence against a cGK-mediated activation of MAP kinases and  $\alpha II_b\beta 3$ integrin by vWF was reported by Marshall *et al.* (2004). They demonstrated a critical role of Src kinases but not MAP kinases in the vWF-dependent activation and also confirmed the inhibitory role for cGMP-elevating agents in regulating this process.

### cGMP AND INSULIN

The above comments summarized some current evidence that the NO/cGMP/cGK signaling pathway is important in the control of many cellular functions. VSMCs are the major constituents of blood vessel walls and are responsible for the maintenance of vascular tone. These cells also contribute to the pathogenesis of type 2 diabetes, hypertension, and cardiovascular diseases (Cohen & Vanhoutte, 1995; Sowers & Epstein, 1995; Sowers et al., 2001; Gewaltig & Kojda, 2002). In these diseases increased contractility of VSMCs, an abnormal vascular tone and defective vasorelaxation are the earliest abnormalities observed. In this section we will therefore briefly review the relationship of the NO/cGMP/cGK signaling pathway and type 2 diabetes.

Insulin can inhibit VSMC contraction, migration and growth in the normal vasculature (Somlyo & Somlyo, 1994; Hsueh & Law, 1999). Failure of this function in insulin-resistance may contribute to enhanced atherosclerosis/restenosis in these clinical cases. Studies with human subjects indicate that hypertension and type 2 diabetes are associated with an impaired vasodilatory response to acetylcholine and SNP, suggesting impaired VSMC responsiveness to NO (Steinberg *et al.*, 1996). A relationship of impaired endothelium-dependent relaxation and expression levels of NOS in diabetic mice was shown (Gunnett *et al.*, 2003).

Recent studies demonstrated that insulin induces relaxation of VSMC via stimulation of

myosin phophatase and inhibition of Rho kinase activity (Sandu et al., 2000; Begum et al., 2000). Further data revealed that insulin inhibits Rho signaling by altering posttranslational modification of RhoA via nitric oxide/cGMP signaling pathway causing myosin-bound phosphatase (MBP) activation, actin cytoskeletal disorganization and vasodilatation (Begum et al., 2000). These effects may contribute to the well-known vasodilator actions of insulin. Therefore, an evaluation of platelet function during development of diabetes is of major interest. Previous studies (Tschoepe *et al.*, 1993) showed that platelets in diabetic patients are larger than those of healthy patients. In addition, platelets of diabetics are characterized by an enhanced activation and aggregation in response to different stimuli in vitro. Also, recently an altered platelet NO level and generation was observed in platelets from healthy donors after insulin-treatment (McKendrick et al., 1998; Fleming et al., 2003). However, the underlying mechanisms and their contribution to the development of cardiovascular diseases have not as yet been established.

Given the importance of the NO/cGMP/ cGK signaling pathway for the cardiovascular system and effects on platelet function as described above, an understanding of the exact mechanism of insulin function and the pathophysiology of the vascular complications associated with diabetes promises to be a good setting-out point for achieving more effective therapeutic tools.

Although much progress has been made in recent years regarding the NO/cGMP/cGK signaling pathway and the effects of different diseases, functional *in vivo* data is often lacking. Hopefully over the next few years progress in molecular and biophysical approaches will give us a clearer picture of the function and regulation of cGK, allowing many important questions to be addressed and also to see profound advancement of further therapies in treating vascular signaling.

### REFERENCES

- Aktas B, Utz A, Hoenig-Liedl P, Walter U, Geiger J. (2003) Dipyridamole enhances NO/cGMP-mediated vasodilator-stimulated phosphoprotein phosphorylation and signaling in human platelets: *in vitro* and *in vivo/ex vivo* studies. *Stroke.*; 34: 764-9.
- Ammendola A, Geiselhoringer A, Hofmann F, Schlossmann J. (2001) Molecular determinants of the interaction between the inositol 1,4,5-trisphosphate receptor-associated cGMP kinase substrate (IRAG) and cGMP kinase Iβ. J Biol Chem.; 276: 24153-9.
- Begum N, Duddy N, Sandu O, Reinzie J, Ragolia L. (2000) Regulation of myosin-bound protein phosphatase by insulin in vascular smooth muscle cells: evaluation of the role of Rho kinase and phosphatidylinositol-3-kinase-dependent signaling pathways. *Mol Endocrinol.*; 14: 1365-76.
- Bhatt DL, Topol EJ. (2003) Scientific and therapeutic advances in antiplatelet therapy. Nat Rev Drug Discov.; 2: 15-28.
- Cohen RA, Vanhoutte PM. (1995) Endothelium-dependent hyperpolarization. Beyond nitric oxide and cyclic GMP. *Circulation.*; 92: 3337-49.
- Etter EF, Eto M, Wardle RL, Brautigan DL, Murphy RA. (2001) Activation of myosin light chain phosphatase in intact arterial smooth muscle during nitric oxide-induced relaxation. J Biol Chem.; 276: 34681-5.
- Fleming I, Schulz C, Fichtlscherer B, Kemp BE, Fisslthaler B, Busse R. (2003) AMP-activated protein kinase (AMPK) regulates the insulin-induced activation of the nitric oxide synthase in human platelets. *Thromb Haemost.*; **90**: 863-71.
- Fukao M, Mason HS, Britton FC, Kenyon JL, Horowitz B, Keef KD. (1999) Cyclic GMP-dependent protein kinase activates cloned BKCa channels expressed in mammalian cells by direct phosphorylation at serine 1072. J Biol Chem.; 274: 10927-35.
- Furchgott RF, Zawadzki JV. (1980) The obligatory role of endothelial cells in the relaxation

of arterial smooth muscle by acetylcholine. *Nature*.; **288**: 373–6.

- Gambaryan S, Geiger J, Schwarz UR, Butt E, Begonja A, Obergfell A, Walter U. (2004) Potent inhibition of human platelets by cGMP analogs independent of cGMP-dependent protein kinase. *Blood.*; 103: 2593-600.
- Gewaltig MT, Kojda G. (2002) Vasoprotection by nitric oxide: mechanisms and therapeutic potential. *Cardiovasc Res.*; **55**: 250-60.
- Gunnett CA, Heistad DD, Faraci FM. (2003) Gene-targeted mice reveal a critical role for inducible nitric oxide synthase in vascular dysfunction during diabetes. *Stroke.*; 34: 2970-4.
- Haslam RJ, Davidson MM, Davies T, Lynham JA, McClenaghan MD. (1978) Regulation of blood platelet function by cyclic nucleotides. Adv Cyclic Nucleotide Res.; 9: 533-52.
- Hedlund P, Aszodi A, Pfeifer A, Alm P,
  Hofmann F, Ahmad M, Fassler R, Andersson KE. (2000) Erectile dysfunction in cyclic
  GMP-dependent kinase I-deficient mice. *Proc* Natl Acad Sci USA.; 97: 2349-54.
- Hofmann F, Ammendola A, Schlossmann J. (2000) Rising behind NO: cGMP-dependent protein kinases. J Cell Sci.; 113: 1671-6.
- Hsueh WA, Law RE. (1999) Insulin signaling in the arterial wall. Am J Cardiol.; 84: 21J-4J.
- Hu H, Hjemdahl P, Li N. (2002) Effects of insulin on platelet and leukocyte activity in whole blood. *Thromb Res.*; 107: 209-15.
- Ignarro LJ, Buga GM, Wood KS, Byrns RE, Chaudhuri G. (1987) Endothelium-derived relaxing factor produced and released from artery and vein is nitric oxide. *Proc Natl Acad Sci USA*.; 84: 9265-9.
- Katsuki S, Arnold W, Mittal C, Murad F. (1977) Stimulation of guanylate cyclase by sodium nitroprusside nitroglycerin and nitric oxide in various tissue preparations and comparison to the effects of sodium azide and hydroxylamine. J Cyclic Nucleotide Res.; **3**: 23-35.
- Li Z, Ajdic J, Eigenthaler M, Du X. (2003a) A predominant role for cAMP-dependent pro-

tein kinase in the cGMP-induced phosphorylation of vasodilator-stimulated phosphoprotein and platelet inhibition in humans. *Blood.*; **101**: 4423-9.

- Li Z, Xi Z, Gu M, Feil R, Ye RD, Eigenthaler M, Hofmann F, Du X. (2003b) A Stimulatory role for cGMP-dependent protein kinase in platelet activation. *Cell.*; **112**: 77-86.
- Lincoln TM, Dey N, Sellak H. (2001) Invited review: cGMP-dependent protein kinase signaling mechanisms in smooth muscle: from the regulation of tone to gene expression. J Appl Physiol.; 91: 1421-30.
- Lohmann SM, Vaandrager AB, Smolenski A, Walter U, De Jonge HR. (1997) Distinct and specific functions of cGMP-dependent protein kinases. *Trends Biochem Sci.*; 22: 307-12.
- Marshall SJ, Senis YA, Auger JM, Feil R, Hofmann F, Salmon G, Peterson JT, Burslem F, Watson SP. (2004) GPIb-dependent platelet activation is dependent on Src kinases but not MAP kinase or cGMP-dependent kinase. *Blood.*; 103: 2601-9.
- Massberg S, Sausbier M, Klatt P, Bauer M, Pfeifer A, Siess W, Fassler R, Ruth P, Krombach F, Hofmann F. (1999) Increased adhesion and aggregation of platelets lacking cyclic guanosine 3',5'-monophosphate kinase I. J Exp Med.; 189: 1255-64.
- McKendrick JD, Salas E, Dube GP, Murat J, Russell JC, Radomski MW. (1998) Inhibition of nitric oxide generation unmasks vascular dysfunction in insulin-resistant obese JCR:LA-cp rats. *Br J Pharmacol.*; **124**: 361-9.
- Munzel T, Feil R, Mulsch A, Lohmann SM, Hofmann F, Walter U. (2003) Physiology and pathophysiology of vascular signaling controlled by guanosine 3'5'-cyclic monophosphate-dependent protein kinase [corrected]. *Circulation.*; **108**: 2172-83.
- Nolte C, Eigenthaler M, Horstrup K, Honig-Liedl P, Walter U. (1994) Synergistic phosphorylation of the focal adhesion-associated vasodilator-stimulated phosphoprotein in intact human platelets in response to

cGMP- and cAMP-elevating platelet inhibitors. *Biochem Pharmacol.*; **48**: 1569–75.

- Ny L, Pfeifer A, Aszodi A, Ahmad M, Alm P, Hedlund P, Fassler R, Andersson KE. (2000) Impaired relaxation of stomach smooth muscle in mice lacking cyclic GMP-dependent protein kinase I. *Br J Pharmacol.*; **129**: 395-401.
- Palmer RM, Ferrige AG, Moncada S. (1987) Nitric oxide release accounts for the biological activity of endothelium-derived relaxing factor. *Nature.*; **327**: 524-6.
- Pfeifer A, Aszodi A, Seidler U, Ruth P, Hofmann F, Fassler R. (1996) Intestinal secretory defects and dwarfism in mice lacking cGMP-dependent protein kinase II. *Science.*; 274: 2082-6.
- Pfeifer A, Klatt P, Massberg S, Ny L, Sausbier M, Hirneiss C, Wang GX, Korth M, Aszodi A, Andersson KE, Krombach F, Mayerhofer A, Ruth P, Fassler R, Hofmann F. (1998) Defective smooth muscle regulation in cGMP kinase I-deficient mice. *EMBO J.*; 17: 3045-51.
- Pfitzer G. (2001) Invited review: regulation of myosin phosphorylation in smooth muscle. J Appl Physiol.; 91: 497–503.
- Ruth P. (1999) Cyclic GMP-dependent protein kinases: understanding *in vivo* functions by gene targeting. *Pharmacol Ther.*; 82: 355-72.
- Sandu OA, Ragolia L, Begum N. (2000) Diabetes in the Goto-Kakizaki rat is accompanied by impaired insulin-mediated myosin-bound phosphatase activation and vascular smooth muscle cell relaxation. *Diabetes.*; 49: 2178-89.
- Sauzeau V, Le Jeune H, Cario-Toumaniantz C, Smolenski A, Lohmann SM, Bertoglio J, Chardin P, Pacaud P, Loirand G. (2000) Cyclic GMP-dependent protein kinase signaling pathway inhibits RhoA-induced Ca<sup>2+</sup> sensitization of contraction in vascular smooth muscle. J Biol Chem.; 275: 21722-9.
- Schlossmann J, Ammendola A, Ashman K, Zong X, Huber A, Neubauer G, Wang GX, Allescher HD, Korth M, Wilm M, Hofmann F, Ruth P. (2000) Regulation of intracellular

calcium by a signalling complex of IRAG IP3 receptor and cGMP kinase I $\beta$ . *Nature.*; **404**: 197–201.

- Schultz KD, Schultz K, Schultz G. (1977) Sodium nitroprusside and other smooth muscle-relaxants increase cyclic GMP levels in rat ductus deferens. *Nature.*; **265**: 750-1.
- Schwarz UR, Walter U, Eigenthaler M. (2001) Taming platelets with cyclic nucleotides. *Biochem Pharmacol.*; 62: 1153-61.
- Somlyo AP, Somlyo AV. (1994) Signal transduction and regulation in smooth muscle. *Nature.*; **372**: 231-6.
- Sowers JR, Epstein M. (1995) Diabetes mellitus and associated hypertension vascular disease and nephropathy. An update. *Hypertension*.; **26**: 869-79.
- Sowers JR, Epstein M, Frohlich ED. (2001) Diabetes hypertension and cardiovascular disease: an update. *Hypertension.*; **37**: 1053-9.
- Steinberg HO, Chaker H, Leaming R, Johnson A, Brechtel G, Baron AD. (1996) Obesity/insulin resistance is associated with endothelial dysfunction. Implications for the syndrome

of insulin resistance. J Clin Invest.; 97: 2601–10.

- Tschoepe D, Roesen P, Schwippert B, Gries FA. (1993) Platelets in diabetes: the role in the hemostatic regulation in atherosclerosis. Semin Thromb Hemost.; 19: 122-8.
- Vaandrager AB, Bot AG, Ruth P, Pfeifer A, Hofmann F, De Jonge HR. (2000) Differential role of cyclic GMP-dependent protein kinase II in ion transport in murine small intestine and colon. *Gastroenterology.*; 118: 108-14.
- von der Leyen HE, Dzau VJ. (2001) Therapeutic potential of nitric oxide synthase gene manipulation. *Circulation.*; **103**: 2760-5.
- Wagner C, Pfeifer A, Ruth P, Hofmann F, Kurtz A. (1998) Role of cGMP-kinase II in the control of renin secretion and renin expression. *J Clin Invest.*; **102**: 1576–82.
- Young DV, Serebryanik D, Janero DR, Tam SW. (2000) Suppression of proliferation of human coronary artery smooth muscle cells by the nitric oxide donor S-nitrosoglutathione is cGMP-independent. Mol Cell Biol Res Commun.; 4: 32-36.