Rituximab therapy for cytomegalovirus-associated autoimmune hemolytic anemia in a liver-transplant recipient

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Cytomegalovirus (CMV)-associated auto-immune hemolytic anemia (AIHA) occurs mostly in infants [1] and in HIV-infected adults [2]. We herein report the first case to occur in a solid-organ-transplant recipient.

A CMV seronegative liver-transplant (LT) recipient was referred to our unit 1 year post-transplantation with fatigue, dyspnea, and icterus. Immunosuppression relied on tacrolimus/mycophenolate mofetil/prednisone. He received valganciclovir prophylaxis until 2 months before admission

(CMV seropositive donor). Physical examination showed pallor and conjunctival icterus. Blood samples revealed acute anemia (hemoglobin 6 g/dl), thrombopenia (120 000/mm³), hepatitis with slight cytolysis (transaminase levels 1.5 N), and cholestasis (γ -glutamyl transferase 4 N). Total/unconjugated bilirubin levels were 78/53 μ mol/l. Reticulocyte count (311 000/mm³), elevated lacto-dehydrogenase (LDH; 4 N), and low haptoglobin (<0.06 g/l) levels defined hemolytic anemia. Toxic or

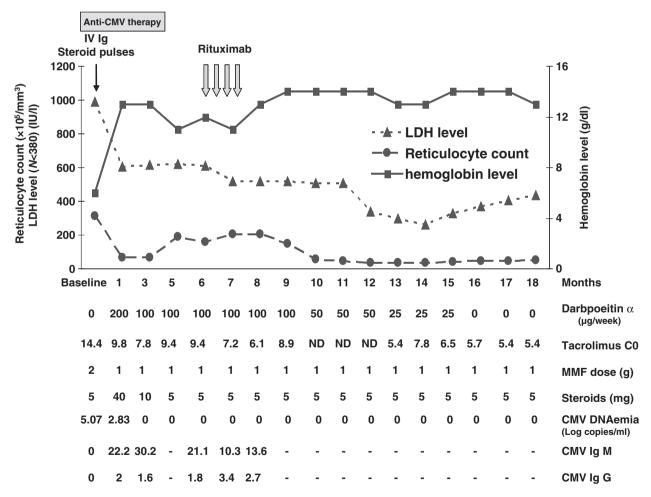


Figure 1 Outcome of hemoglobin and lacto-dehydrogenase (LDH) levels, as well as reticulocyte count after cytomegalovirus-associated auto-immune hemolytic anemia.

immuno-allergic hemolytic anemia was excluded (sole co-medication: insulin). Blood smears showed no morphological abnormalities in erythrocytes, with absence of schistocytes. Identification of cold agglutinins and a positive Coombs' test for complement fraction C3d (titer > 1/8) made the diagnosis of AIHA in a setting of primary CMV-infection (CMV DNAemia: 5.07 log/ml whole blood; CMV antibody [IgM]). Other virological tests, including HIV, human Herpes viruses 6 and 8, Epstein–Barr virus, and hepatitis A, B, C, and E viruses, were all negative. Immunological testing showed low titers of antinuclear antibodies (1/160) and rheumatoid factor (100 UI/ml). There was no evidence of monoclonal proliferation nor of cryoglobulinemia.

Antiviral treatment was 3 weeks of intravenous ganciclovir (10 mg/kg/day) followed by oral valganciclovir (900 mg/day) for another 3 months. Under this treatment, CMV DNAemia and CMV IgM antibodies became negative within 2 and 4 weeks, respectively and remained so until last follow-up. Anti-CMV IgG became positive 21 days after starting ganciclovir at high titer which remained elevated until last follow-up. AIHA was treated by three pulses of methylprednisolone (2 mg/kg/ day), followed by oral prednisolone (1 mg/kg/day) for a month, before steroids were progressively tapered. Concomitantly, he received 0.5 g/kg/day of intravenous immunoglobulins (IVIg) for the first 4 days, a blood transfusion, and darbopoietin-α (200 µg/week). Nevertheless, the hemolytic process continued; thus, at 6 months after initial AIHA diagnosis, 100 µg/week of darbopoietin-α was still necessary to maintain stable hemoglobin levels. Therefore, we implemented rituximab therapy (four weekly courses of 375 mg/m² each). Three months after the last rituximab infusion, although haptoglobin levels remained low, LDH and reticulocytes were markedly reduced and hemoglobin was stable. Cold agglutinins were hardly positive at last follow-up. Darbopoetin-α was progressively stopped. Complete remission of AIHA has been maintained after 1 year follow-up (Fig. 1).

The pathophysiology of CMV-associated AIHA remains unclear [3]. CMV infection might result in auto-reactive

B-lymphocyte clones. Cross-reactivity between viral epitopes and erythrocytic antigens is also possible [4].

Conventional treatment of AIHA relies on immunosuppression (steroids and/or IVIg) with or without splenectomy [5]. However, recently, rituximab has been used with good results in some cases [6]. In CMV-related AIHA, patients have been additionally treated with ganciclovir, or with anti-CMV globulins.

In conclusion, we report the first case of CMV-associated AIHA in a LT patient. Complete remission was obtained with rituximab infusions.

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